

Systematic evaluation of 1-chlorobutane for liquid-liquid extraction of drugs

U. Demme^{1*}, J. Becker², H. Bussemas³, T. Daldrup⁴, F. Erdmann⁵, M. Erkens⁶, P.X. Iten⁷, H. Käferstein⁸, K.J. Lusthof⁹, H.J. Mager¹⁰, L.v. Meyer¹¹, A. Reiter¹², G. Rochholz¹³, A. Schmoldt¹⁴, E. Schneider¹⁵, H.W. Schütz¹³, T. Stimpfl¹⁶, F. Tarbah¹⁷, J. Teske¹⁸, W. Vycudilik¹⁶, J.P. Weller¹⁸, W. Weinmann¹⁹



*On behalf of the "Workgroup Extraction" of the GTFCh (Society of Toxicological and Forensic Chemistry, Germany)

Institutes of Forensic Medicine of ¹Jena, ²Mainz, ³Duesseldorf, ⁵Giessen, ⁷Zuerich (CH), ⁸Cologne, ¹⁰Wuerzburg, ¹¹Munich, ¹²Luebeck, ¹³Kiel, ¹⁴Hamburg, ¹⁶Vienna (A), ¹⁸Hannover, ¹⁹Freiburg and ³Praxis Labormedizin Dortmund, ⁶Clin.-Chem. Central Laboratory, Aachen, ⁹Nat. For. Inst., Den Haag (NL), ¹⁵LKA Baden-Wuerttemberg, Stuttgart ¹⁷Dubai Police Dept.

Introduction

In systematic toxicological analysis (STA), chromatographic methods are widely used for the detection of drugs and other organic toxic substances in biological materials, such as blood, plasma, urine, hair and tissue samples. In most cases the applied analytical methods like gas chromatography-mass spectrometry (GC/MS), high performance liquid chromatography with diode-array detection (HPLC/DAD), HPLC/mass spectrometry (LC/MS), or thin-layer chromatography with remission spectroscopy (TLC/UV), require sample preparation by extraction prior to analysis.

The most widely used procedures for isolation of drugs from biological materials are Liquid-Liquid-Extraction (LLE) and Solid-Phase-Extraction (SPE). The basis of LLE is the application of organic, non-water-miscible solvents at varying pH-values, including basic or acidic back-extraction; sometimes after immobilization onto diatomaceous earth, silica gel, celite or sodium sulfate. Verified information about the extractability of drugs provides the basis for method development in LLE, making the realization of LLE faster in routine analyses and thus providing a cheaper alternative to SPE. The **theoretical basis of LLE** is rather simple. The extraction yield only depends on two constants, as shown in the equation for basic compounds. K_D is the distribution constant (Nernst's law of distribution) and K_B is the dissociation constant of the given basic drug (V_o and V_a stand for the volumes of the organic and aqueous phase, Y^o is the extraction yield in the organic phase). The dissociation constants of many drugs are known [4]. If distribution constants are known too, it is possible to determine the extraction yield for a given condition (pH-value (C_{H^+}) and phase volumes). This is only valid for pure aqueous phases and not in the case of biological materials. But the exact determination of K_D -values is rather time consuming, it requires several measurements of extraction yields at different pH-values, near the point of equivalence (Y^o between 0.1 and 0.9) to minimize the error during determination.

$$Y^o = \frac{K_D \times V_o}{K_D \times V_o + V_a (1 + 10^{14} \times K_B \times c_{H^+})}$$

The **aim** of this study was to obtain data on the extractability of many toxicological and therapeutic relevant drugs using 1-chlorobutane. This solvent showed the cleanest chromatograms with regard to interfering peaks in relation to an external standard; in a pilot study 1-chlorobutane was compared to 7 other solvents and solvent mixtures using a pooled serum and GC-MS and HPLC-DAD [1-3]. Numerous publications applied 1-chlorobutane [18-42], also in comparison to SPE [5-17]; but only a few recent papers exist, or they are restricted to distinct classes of drugs. None of these papers contain the newer drugs developed in the last years on the legal and illegal market. For better reproducibility the determinations should be performed for each drug in different laboratories. For these reasons (great number of drugs and measurement in different laboratories) we did not determine the distribution constants, but focussed on the extraction yield for each substance for a distinct pH-value using aqueous buffer solutions. Furthermore, a very simple analytical method was used, based on UV-VIS-spectrometry.

Experimental

Buffer solutions (pH 9) were obtained by dissolving 10 g Na_2HPO_4 (VWR) in one liter of distilled water. 1-chlorobutane and methanol (analytical grade) were obtained from VWR International (Darmstadt, Germany). Drugs were purchased as pure compounds or as salts from chemical suppliers (Sigma Deisenhofen/Germany and others) or from the pharmaceutical manufacturers and were first dissolved in methanol to achieve a concentration of 1 mg/mL. In some instances the pure drug was not available; in these cases, a methanolic solution of a pharmaceutical preparation was used.

Measurement of extraction yields: For obtaining reference solutions three different procedures have been used: a) drugs soluble in the organic solvent were dissolved in 1-chlorobutane in a concentration of 10 or 20 mg/L, depending on the extinction coefficient; b) water-soluble drugs were dissolved in phosphate buffer (pH 9) and c) salts of basic drugs were transferred to their free bases by extraction with 1-chlorobutane at pH 9 from aqueous solution. From the resulting reference solutions UV-spectra were measured to obtain a reference value (A^{pre}). LLE was then performed by adding 3.0 ml of phosphate buffer (pH 9) or - in case of aqueous reference solutions - 3.0 ml 1-chlorobutane, to 3.0 ml of reference solution. In any case, LLE was performed using equal volumes of aqueous and organic phases (v/v : 1:1). The samples were vortex-mixed for 1 min and centrifuged to separate the phases. Either the organic phase (for unpolars compounds) or the aqueous phase (for water-soluble compounds) was measured by UV-spectroscopy (A^{post}). Each LLE was performed twice.

The extraction efficiency (yield) Y^o in the organic phase was determined by the ratio of the absorbance obtained after and before the extraction:

UV-measurement using the **organic** phase: $Y^o = \frac{A_o^{post}}{A_o^{pre}}$

UV-measurement using the **aqueous** phase: $Y^o = \frac{A_a^{pre} - A_a^{post}}{A_a^{pre}}$

A_o^{post} , A_o^{pre} absorbance of the organic phase (o) pre- and post-extraction.

A_a^{post} , A_a^{pre} absorbance of the aqueous phase (a) pre- and post-extraction.

For a minor number of substances, which did not show a typical UV-spectrum in sufficient intensity, quantitation was performed by GC/MS using calibration with external standards.

Results

The extraction yields for 333 compounds of clinical or forensic interest have been determined, including psychopharmaceuticals and narcotics (e.g., benzodiazepines, tricyclic antidepressants, phenothiazines, thioxanthenes, butyrophenones), analgesics, antiepileptics, cardiovascular drugs (e.g., β -blockers, Ca-antagonists), illicit drugs and pesticides.

Extraction yields are listed in alphabetical order (in the table on this poster only drugs with the initial letter A and B with their yield in the organic phase Y^o can be shown). Average values of two to four laboratories are given (one reference laboratory and one to three other laboratories). Since each analyte was extracted twice in each laboratory, the extraction efficiencies obtained by 4 to 8 single measurements were averaged, and the average value is given with one digit behind the decimal point, except in those cases with very poor or very good extraction yields ($Y^o < 0.2$ or $Y^o > 0.8$), when the average value is given with a precision of 0.05. In the last column, references for the extraction with 1-chlorobutane from biological material are given. 228 of 333 drugs (68 %) were extracted by 1-chlorobutane with extraction yields ≥ 80 % ($Y^o \geq 0.8$) from aqueous buffer at pH 9. For 56 compounds, analytical methods have been published using 1-chlorobutane for the extraction of biological material.

However, low extraction efficiencies were obtained, when the drug was not soluble enough in 1-chlorobutane (e.g., morphine, benzoylcegonine, caffeine), and with acidic compounds since the extraction was performed at pH 9 (e.g. diclofenac, indomethacin, phenobarbitone).

The complete table and list of references is available via web: www.gtfch.org/chlorobutaneextraction.pdf

Discussion

The presented list of extraction yields of 333 compounds allows the analyst an easy estimation of the extractability of common drugs using 1-chlorobutane. A high recovery from the aqueous solution is a "necessary but not sufficient" condition for the extraction of biological matrices such as serum, blood or urine; in other words, it is not necessary to test the extraction of a given drug from serum with 1-chlorobutane, if recovery from the aqueous solution is low or even zero. Therefore, and due to a large number of compounds of interest in forensic toxicology, extraction from the aqueous solution (phosphate buffer, pH 9) was investigated. 68 % of the tested drugs were extractable with high yields (> 80%) at pH 9 using 1-chlorobutane. For 56 compounds with high recovery from buffer solution, a successful extraction by 1-chlorobutane from serum was confirmed in the literature and a comprehensive bibliography is given (see Table). In these cases, extraction yields were similar to those found for aqueous solutions in our work. Therefore, in cases of target compound analysis (e.g. detection of poisoning with - or misuse of - a known drug and therapeutic drug monitoring), the table allows the analyst to decide whether an extraction with 1-chlorobutane seems useful in this particular case. For systematic toxicological analysis (STA) it should be taken into account that only 228 of the listed compounds could be extracted with a sufficient extraction yield, and about 32 % of the 333 compounds tested showed poor extraction yields. Therefore, a more polar solvent, mixture of polar solvents or even another methodology (e.g. SPE) should be applied for STA. For the use of a more polar solvent (e.g. ethylacetate) the table can also be very useful, because it can be assumed that drugs with a medium recovery rate in 1-chlorobutane are extractable in this solvent. Future work will include extraction yields for the extraction of the tested compounds from serum samples with 1-chlorobutane.

N°	drug	Y^o	Ref.
24	Acebutolol	0.05	
25	Adenosin	0	
26	Ajmaline	0.5	
27	Alfentanil	1	
28	Alimemazine	1	
29	Alprazolam	0.95	[43-45]
30	Alprenolol	1	
31	Amantadine	0.5	
32	Amfebutammon	1	
33	Amfepramon	1	
34	Amfetaminil	1	
35	Amiodarone	0.95	
36	Amisulprid	0.6	
37	Amitriptyline	1	[46]
38	Amitriptyline-Oxid	0.1	
39	Amlodipine	1	
40	Amphetamine	0.5	[47-55]
41	Apomorphin	0.8	
42	Aprindin	0.95	
43	Articain	1	
44	Atenolol	0	
45	Atropine	0.6	
46	Azathioprine	0	
47	Azinphos-ethyl	1	
48	Azinphos-methyl	1	
49	Benperidol	1	
50	Benserazid	0	
51	Benzatropin	1	
52	Benzoylcegonine	0	
53	Betaxolol	1	
54	Biperidene	1	
55	Bisacodyl	0.7	
56	Bisoprolol	0.9	
57	Bromazepam	0.9	
58	Bromocriptin	1	
59	Bromophos-ethyl	1	

(cont.: see www.gtfch.org/chlorobutaneextraction.pdf)

Table: Extraction yields using 1-chlorobutane from aqueous solution (pH 9).

No.	Compound	Y°	References
1	10-Hydroxy-Carbazepin	0	
2	2-(2,3-methylenedioxyphenyl)butan-1-amin	0.8	
3	2-(2,3-methylenedioxyphenyl)propan-1-amin	0.7	
4	2-(3,4-methylenedioxyphenyl)2-methylpropan-1-amin	0.9	
5	2-(3,4-methylenedioxyphenyl)butan-1-amin	0.8	
6	2-(3,4-methylenedioxyphenyl)propan-1-amin	0.5	
7	2,3 – MDA	0.6	
8	2,3 – MDMA	0.9	
9	2,3-Methylenedioxy-N-methylphenethylamin	0.6	
10	2,3-Methylenedioxy-N-phenethylamin	0.3	
11	2,4,5-Trimethoxy-amphetamin (TMA-2)	0.2	
12	2,4,6-Trimethoxy-amphetamin (TMA-6)	0.5	
13	2,5-Dimethoxy-4-brom-phenethylamin (2-CB)	0.8	
14	2,5-Dimethoxy-4-methyl-amphetamin (DOM)	0.8	
15	2,5-Dimethoxy-4-methyl-phenethylamin (2-CD)	0.5	
16	2,5-Dimethoxy-phenethylamin (2-C-A)	0.3	
17	3-(2,3-methylenedioxyphenyl)pentan-2-amin	0.95	
19	3,4-Methylenedioxy-N-methylphenethylamin	0.5	
20	3,4-Methylenedioxy-N-phenethylamin	0.3	
21	4-Methoxy-2-pyrrolidinopropiophenon	1	
22	4-Methoxy-amphetamin (PMA)	0.5	
23	4-Methyl-2- pyrrolidinopropiophenon (MPPP)	1	
24	Acebutolol	0.05	
25	Adenosin	0	
26	Ajmaline	0.5	
27	Alfentanil	1	
28	Alimemazine	1	
29	Alprazolam	0.95	(43 - 45)
30	Alprenolol	1	
31	Amantadine	0.5	
32	Amfebutamon	1	
33	Amfepramon	1	
34	Amfetaminil	1	
35	Amiodarone	0.95	
36	Amisulprid	0.6	
37	Amitriptyline	1	(46)
38	Amitriptylin-Oxid	0.1	
39	Amlodipine	1	
40	Amphetamine	0.5	(47 - 55)
41	Apomorphin	0.8	
42	Aprindin	0.95	
43	Articain	1	
44	Atenolol	0	
45	Atropine	0.6	
46	Azathioprine	0	
47	Azinphos-ethyl	1	
48	Azinphos-methyl	1	
49	Benperidol	1	
50	Benserazid	0	
51	Benzatropin	1	
52	Benzoylcegonine	0	

No.	Compound	Y°	References
53	Betaxolol	1	
54	Biperidene	1	
55	Bisacodyl	0.7	
56	Bisoprolol	0.9	
57	Bromazepam	0.9	
58	Bromocriptin	1	
59	Bromophos-ethyl	1	
60	Bromophos-methyl	1	
61	Bromperidol	1	
62	Brotizolam	1	
63	Budipine	1	
64	Bupivacaine	1	
65	Bupranolol	1	
66	Buprenorphine	1	
67	Buspiron	0.95	(56)
68	Caffein	0.3	
69	Carazolol	0.9	
70	Carbamazepine	0.95	
71	Carbamazepine-Epoxide	0.6	
72	Carbidopa	0	
73	Carteolol	0.06	
74	Carvedilol	1	
75	Cathin	0.07	
76	Celiprolol	0.1	
77	Chlordiazepoxide	0.95	
78	Chlormezanone	0.9	
79	Chloroquine	0.95	
80	Chlorovinphos	1	
81	Chlorpromazine	1	
82	Chlorprothixene	1	
83	Citalopram	1	(57)
84	Clobazam	1	
85	Clobutinol	1	
86	Clomethiazol	1	
87	Clomipramine	1	
88	Clonazepam	1	(58, 59)
89	Clopidamid	0.06	
90	Clopendixol	1	
91	Clotiazepam	1	
92	Clozapine	1	(60)
93	Cocaine	1	(61- 64)
94	Codeine	0.8	(65)
95	Colchicin	0.13	
96	Cotinine	0.1	(66)
97	Cyamemazin	1	
98	Deanol	0.4	
99	Demelverine	1	
100	Desipramine	1	(46)
101	Detajmium	0.9	
102	Dialifos	1	
103	Diazepam	0.95	(59,65,67)
104	Dibenzepine	1	
105	Dichlorovos	1	
106	Diclofenac	0.2	

No.	Compound	Y°	References
107	Dihydrocodeine	0.7	
108	Dihydroergocryptin	1	
109	Diltiazem	1	(68)
110	Dimethoate	0.7	
111	Dimetindene	0.98	
112	Diphenhydramine	1	(65)
113	Disopyramide	0.8	(69)
114	Dosulepine	1	
115	Doxazosine	0.95	
116	Doxepine	1	(46,70,71)
117	Doxylamine	1	
118	Droperidol	0.9	
119	Enalapril	0	
120	Entacapon	0	
121	Ephedrine	0.2	
122	Esmolol	0.8	
123	Ethosuximide	0.2	
124	Etomidate	1	
125	Felodipin	0.95	
126	Fenethylline	0.9	
127	Fenofos	1	
128	Fentanyl	1	(72 - 74)
129	Fenthion	1	
130	Flecainide	0.95	(75, 76)
131	Fluconazol	0.1	
132	Flumazenil	0.8	
133	Flunitrazepam	1	(59, 67)
134	Fluoxetine	0.8	(77, 78)
135	Flupentixol	1	
136	Fluphenazine	1	
137	Flupirtine	1	
138	Flurazepam	0.95	
139	Fluspirilen	0.9	
140	Fluvoxamine	0.8	
141	Furosemide	0	
142	Gabapentin	0	
143	Gallopamil	1	
144	Gamma-hydroxybutyric acid	0	
145	Glibenclamid	0.2	
146	Glutethimide	1	
147	Haloperidol	1	(79)
149	Heptenophos	0.8	
148	Hydrochlorothiazide	0	
150	Hydromorphon	0.1	
151	Hydroxyzin	1	
152	Ibuprofen	0	
153	Imipramine	1	(46)
154	Indomethacin	0	
155	Ipratropium	0	
156	Isofenphos	0.95	
157	Kavain	0.7	
158	Ketamine	1	
159	Lamotrigine	0.17	(80)
160	Levetiracetam	0	

No.	Compound	Y°	References
161	Levodopa	0	
162	Levomepromazine	1	
163	Lidocaine	1	
164	Lisinopril	0	
165	Lofepramin	0.9	
166	Loprazolam	1	
167	Lorazepam	0.85	(44)
168	Lormetazepam	1	
169	Loxapine	1	
170	LSD	0.95	(81, 82)
171	Maprotiline	1	
172	MBDB	1	
173	MDA	0.6	(55,83)
174	MDEA	0.9	(55)
175	MDMA	0.7	(55, 83)
176	Medazepam	1	
177	Mefenorex	0.95	
178	Melperone	1	
179	Mepindolol	0.4	
180	Mepivacain	1	
181	Meprobamat	0.1	
182	Meptazinol	0.9	
183	Mesuximid	1	
184	Metamizol	0.35	
185	Methadone	0.95	(84 - 86)
186	Methamphetamine	0.7	(47,48,50,52 - 54,87)
187	Methaqualone	1	(65,88)
188	Methohexital	0.95	
189	Methylphenidate	0.75	
190	Metixen	1	
191	Metoclopramide	0.9	
192	Metoprolol	0.8	(89)
193	Mevinphos	0.9	
194	Mexiletine	0.9	(90, 91)
195	Mianserine	1	
196	Midazolam	0.9	(59,92, 93)
197	Mirtazapine	0.9	(94)
198	Moclobemide	0.9	(95)
199	Modafinil	0.35	
200	Morphine	0	
201	MTA	0.6	
202	N-(1-Phenylcyclohexyl)-2-methoxy-ethylamin (PCMEA)	1	
203	N-(1-Phenylcyclohexyl)-3-ethoxy-propylamin (PCEPA)	1	
204	N-(1-Phenylcyclohexyl)-3-methoxy-propylamin (PCMPA)	1	
205	N-(1-Phenylcyclohexyl-1)-propylamin	1	
206	N,N-Diethyltryptamin	0.95	
207	Nadolol	0	
208	Nalbuphine	0.8	
209	Nalorphin	0.3	
210	Naloxon	0.9	

No.	Compound	Y°	References
211	Nefazodon	1	
212	Nefopam	0.95	
213	Nicotine	0.9	(66)
214	Nifedipin	1	
215	Nikotinamide	0	
216	Nimodipin	1	
217	Nisoldipin	1	
218	Nitrazepam	1	
219	N-Methyl-4-methoxy-amphetamin (PMMA)	0.7	
220	Nordazepam	0.95	(44, 67)
221	Nortriptyline	1	(46, 96)
222	Noscapine	1	
223	Olanzapin	1	
224	Opipramol	1	
225	Orciprenaline	0	
226	Oxazepam	0.85	(59, 67)
227	Oxcarbazepine	0.9	
228	Oxitriptan	0	
229	Oxprenolol	0.9	
230	Oxycodone	0.95	
231	Paracetamol	0	
232	Paraoxon	0.9	
233	Parathion ethyl	1	
234	Parathion methyl	1	
235	Paroxetine	1	(95, 97)
236	Pemoline	0	
237	Penbutolol	1	
238	Pentazocine	0.8	
239	Pentobarbital	0.2	
240	Pentoxyverine	1	
241	Perazine	1	
242	Perphenazine	1	(98)
243	Pethidine	1	
244	Phenazone	0.35	
245	Phencyclidine	0.9	(99)
246	Phenobarbitone	0.1	
247	Phenolphthalein	0.7	
248	Phenprocoumon	0.2	
249	Phenytoin	0.5	
250	Pholedrin	0	
251	Phosphamidon	0.8	
252	Phoxim	1	
253	Pimozid	1	
254	Pindolol	0.4	
255	Pindolol	0.2	
256	Pipamperone	1	
257	Pirimiphos	1	
258	Piritramid	0.9	
259	Piroxicam	0	
260	Prajmaline	0.9	
261	Pramipexol	0	
262	Prazepam	1	
263	Prilocain	0.95	
264	Primidone	0	

No.	Compound	Y°	References
265	Procain	0.9	
266	Procainamid	0.1	
267	Procyclidin	1	
268	Promazine	1	
269	Promethazine	1	
270	Propafenon	0.95	
271	Propofol	0.95	
272	Propoxyphene	1	
273	Propranolol	1	
274	Propyphenazone	1	
275	Prothipendyl	0.95	
276	Pseudoephedrine	0.2	
277	Quetiapine	1	(100)
278	Quinidine	0.95	
279	Quinine	1	
280	Ranitidine	0	
281	Reboxetine	1	
282	Remifentanil	1	
283	Risperidon	1	
284	Ropivacain	1	
285	Salicylat	0	
286	Scopolamine	0.7	
287	Sertindol	1	
288	Sertraline	1	(101 - 103)
289	Sildenafil	1	
290	Sotalol	0	
291	Strychnine	0.9	(104)
292	Sulfentanil	1	
293	Sulfotep	1	
294	Sulpiride	0	
295	Sultiam	0	
296	Talinolol	0.2	
297	Temazepam	1	(67, 105)
298	Terbufos	1	
299	Tertatolol	1	
300	Tetrazepam	0.95	
301	Theobromin	0	
302	Theophylline	0	
303	Thiopental	0.9	(106)
304	Thioridazine	1	(107)
305	Tiagabin	0.5	
306	Tiapride	0.4	
307	Ticlopidine	1	
308	Tilidine	1	
309	Timolol	0.6	
310	Tocainide	0.3	
311	Tolperison	1	
312	Topiramat	0.2	
313	Tramadol	1	(108)
314	Tranylcypromine	1	(109, 110)
315	Trazodone	1	(111, 112)
316	Triazolam	1	(44, 45, 113)
317	Trichlorophos	1	
318	Trifluperidol	1	

No.	Compound	Y°	References
319	Triflupromazine	1	
320	Trihexyphenidyl	1	
321	Trimipramine	1	
322	Tryptophan	0	
323	Valproinsäure	0.07	
324	Venlafaxine	0.95	(114 - 117)
325	Verapamil	1	(118 - 120)
326	Vigabatrin	0	
327	Viloxazine	0.85	
328	Zaleplon	1	
329	Ziprasidon	1	
330	Zolpidem	1	(121)
331	Zopiclon	0.9	(122)
332	Zotepine	0.95	
333	Zuclopenthixol	1	

References

1. U. Demme, J. Becker, B. Ahrens, H. Bussemas, A. Gräfe, T. Daldrup, L.v.Meyer, K. Padmanaban, A. Reiter, K. Schmidt, A. Schmoltdt, H.W. Schütz and W. Weinmann. Blindwerte bei der Serumextraktion mit Extraktionsmitteln unterschiedlicher Polarität. *Toxichem + Krimtech* **65**: 13-17 (1998)
2. K. Pflieger, H.H. Maurer, A. Weber. Standard extraction for plasma or gastric contents. In: *Mass spectral and GC data of drugs, poisons, pesticides, pollutants and their metabolites*. 2nd Edit., Wiley VCH, Weinheim, New York, Chichester, Brisbane, Singapore, Toronto, Part 4, 5 (2000)
3. K. Pflieger, H.H. Maurer, A. Weber. STA hydrolysis, extraction and microwave-assisted acetylation procedure for urine. In: *Mass spectral and GC data of drugs, poisons, pesticides, pollutants and their metabolites*. 2nd Edit., Wiley VCH, Weinheim, New York, Chichester, Brisbane, Singapore, Toronto, Part 4, 5 (2000)
4. A.C. Moffat, J.V. Jackson, M.S.Moss and B. Widdop. *Clarke's Isolation and Identification of Drugs*. The Pharmaceutical Press, London (1986)
5. X. Chen, J. Wijsbeek, J. Van Veen, J.P. Franke, and R.A. de Zeeuw. Solid phase extraction for the screening of acidic, neutral and basic drugs in plasma using a single column procedure on Bond-Elut Certify. *J. Chromatogr.* **529**: 161-166 (1990)
6. B.K. Logan, C.M. Moore, I.R. Tebbett, and D.T. Stafford. Rapid screening for 100 basic drugs in urine using cation exchange solid-phase extraction

-
- and high performance liquid chromatography with diode array detection. *J. Anal. Toxicol.* **14**: 154-159 (1990)
7. J. Scheurer and C.M. Moore. Solid-phase extraction of drugs from biological tissues – a review. *J. Anal. Toxicol.* **16**: 264-269 (1992)
 8. X. Chen, J.P. Franke, J. Wijsbeek, and R.A. de Zeeuw. Isolation of acidic, neutral and basic drugs from whole blood using a single mixed-mode solid phase extraction column. *J. Anal. Toxicol.* **16**: 351-355 (1992)
 9. X.H. Chen, J. Wijsbeek, J.P. Franke and R.A. de Zeeuw. A single-column procedure on Bond Elut Certify for systematic toxicological analysis of drugs in plasma and urine. *J. Forensic Sci.* **37**: 61-71 (1992)
 10. X.H. Chen, J.P. Franke, K. Ensing, J. Wijsbeek, and R.A. de Zeeuw. Pitfalls and solutions in the development of a fully automated solid-phase extraction method for drug screening purposes in plasma and whole blood. *J. Anal. Toxicol.* **17**: 421-426 (1993)
 11. X.H. Chen, J.P. Franke, J. Wijsbeek, and R.A. de Zeeuw. Determination of basic drugs extracted from biological matrices by means of solid-phase extraction and wide-bore capillary gas chromatography with nitrogen-phosphorous detection. *J. Anal. Toxicol.* **18**: 150-153 (1994)
 12. M.A. Martínez, C. Sánchez de la Torre, E. Almarza. Simultaneous determination of viloxazine, venlafaxine, imipramine, desipramine, sertraline, and amoxapine in whole blood: comparison of two extraction/cleanup procedures for capillary gas chromatography with nitrogen-phosphorus detection. *J. Anal. Toxicol.* **26**: 296-302 (2002)
 13. Z.P. Huang, X.H. Chen, J. Wijsbeek, J.P. Franke, and R.A. de Zeeuw. An enzymic digestion and solid-phase extraction procedure for the screening for acidic, neutral, and basic drugs in liver using gas chromatography for analysis. *J. Anal. Toxicol.* **20**: 248-254 (1996)
 14. S.P. Elliott and K.A. Hale. Applications of an HPLC-DAD drug-screening system based on retention indices and UV spectra. *J. Anal. Toxicol.* **22**: 279-289 (1998)
 15. J.P. Franke and R.A. de Zeeuw. Solid-phase extraction procedures in systematic toxicological analysis. *J. Chromatogr. B* **713**: 51-59 (1998)
 16. T. Soriano, C. Jurado, M. Menéndez and M. Repetto. Improved solid-phase extraction method for systematic toxicological analysis in biological fluids. *J. Anal. Toxicol.* **25**: 137-143 (2001)
 17. M. Rittner, F. Pragst, W.-R. Bork and J. Neumann. Screening method for seventy psychoactive drugs or drug metabolites in serum based on high performance liquid chromatography-electrospray ionization mass spectrometry. *J. Anal. Toxicol.* **25**: 115-124 (2001)
 18. E.H. Foerster and M.F. Mason. Preliminary results on the use of n-butyl chloride as an extractant in a drug screening procedure. *J. Forens. Sci.* **19(1)**: 155-161 (1974)

-
19. R.C. Baselt, C.B. Stewart and S.J. Franch. Toxicological determination of benzodiazepines in biological fluids and tissues by flame-ionization gas chromatography. *J. Anal. Toxicol.* **1**: 10 (1977)
 20. W.O. Pierce, T. C. Lamoreaux, F. M. Urry, L. Kopjak and B.S. Finkle. A new, rapid gas chromatography method for the detection of basic drugs in postmortem blood, using a nitrogen phosphorous detector. Part I. qualitative analysis. *J. Anal. Toxicol.* **2**: 26-31 (1978)
 21. E.H. Foerster, D. Hatchett, and J.C. Garriott. A rapid, comprehensive, screening procedure for basic drugs in blood or tissues by gas chromatography. *J. Anal. Toxicol.* **2**: 50-55 (1978)
 22. T.J. Siek. Effective use of organic solvents to remove drugs from biologic specimens. *Clin. Toxicol.* **13**: 205-230 (1978)
 23. L. Kopjak, B.S. Finkle, T. C. Lamoreaux, W.O. Pierce and F. M. Urry. Rapid gas chromatography method for the detection of basic drugs in postmortem blood, using a nitrogen phosphorous detector. Part II. quantitative analysis. *J. Anal. Toxicol.* **3**: 155-16 (1979)
 24. S.J. Dickson, W.T. Cleary, A.W. Missen, E.A. Queree, and S.M. Shaw. The relative efficiency of drug extraction procedures for enzyme digested livers. *J. Anal. Toxicol.* **4**: 74-77 (1980)
 25. M. Hata, S. Takahashi, K. Matsubara and Y. Fukui. The use of n-Butylchloride as extracting solvent in gas chromatographic analysis for quantitative determination of some basic drugs. *Jap. J. Legal Med.* **34**: 645-650 (1980)
 26. H.W. Peel and B.J. Perrigo. Toxicological analysis of benzodiazepines-type compounds in post-mortem blood by gas chromatography. *J. Anal. Toxicol.* **4**: 105 (1980)
 27. G.W. Hime and L.R. Bednarczyk. Identification of basic drugs in urine by dual fused silica capillary column GC. *J. Anal. Toxicol.* **6**: 247-252 (1982)
 28. J.H. Hebb, Jr., C.R. Crooks, Y.H. Caplan, W.J. Mergner. A method for the determination of therapeutic and toxic concentrations of tricyclic antidepressant drugs in post mortem fluids and tissues. *J. Anal. Toxicol.* **6**: 206-208 (1982)
 29. D.N. Bailey and M. Kelner. Extraction of acidic drugs from water and plasma: study of recovery with five different solvents. *J. Anal. Toxicol.* **8**: 26-28 (1984)
 30. P.D. Whitter and P.L. Cary. A rapid method for the identification of acidic, neutral, and basic drugs in postmortem liver specimens by Toxi-Lab. *J. Anal. Toxicol.* **10**: 68-71 (1986)
 31. V.W. Watts and T.F. Simonick. Screening of basic drugs in biological samples using dual column capillary chromatography and nitrogen phosphorous detectors. *J. Anal. Toxicol.* **10**: 198-204 (1986)

-
32. O.H. Drummer, S. Horomides, S. Koertis, M.L. Syrjanen, and P. Tippet. Capillary gas chromatographic drug screen for use in forensic toxicology. *J. Anal. Toxicol.* **18**: 134-138 (1994)
 33. W.E. Lambert, E. Meyer, and A.P. De Leenheer. Systematic toxicological analysis of basic drugs by gradient elution of an alumina-based HPLC packing material under alkaline conditions. *J. Anal. Toxicol.* **19**: 73-78 (1995)
 34. J.C. Hudson et al. *Can. Soc. Forens. Sci. J.* **28**: 137-152 (1995)
 35. A. Solans, M. Carnicero, R. de la Torre, and J. Segura. Comprehensive screening procedure for detection of stimulants, narcotics, adrenergic drugs, and their metabolites in human urine. *J. Anal. Toxicol.* **19**: 104-114 (1995)
 36. W. Huang and D.E. Moody. Immunoassay detection of benzodiazepines and benzodiazepine metabolites in blood. *J. Anal. Toxicol.* **19**: 333-342(1995)
 37. O.H. Drummer. Methods for the measurement of benzodiazepines in biological samples. *J. Chromatogr. B* **713**: 201-225 (1998)
 38. R.K. Müller. Parameter der flüssig/flüssig-Extraktion toxikologisch relevanter organischer Verbindungen. 1. Mitteilung: Methodik der experimentellen Ermittlung der pH-abhängigen Verteilungsquotienten (Ergebnisse für saure Verbindungen). *Die Pharmazie* **37**: 416-419 (1982)
 39. R.K. Müller. Parameter der flüssig/flüssig-Extraktion toxikologisch relevanter organischer Verbindungen. 2. Mitteilung: Abhängigkeit der Extraktionsausbeute von Verteilungskonstanten, Phasenverhältnis und Stufenzahl (Ergebnisse für neutrale Verbindungen). *Die Pharmazie* **38**: 462-466 (1983)
 40. R.K. Müller. Parameter der flüssig/flüssig-Extraktion toxikologisch relevanter organischer Verbindungen. 3. Mitteilung: Extraktionsausbeute und Fraktionierung (pH-abhängige Verteilungsquotienten für basische Verbindungen). *Die Pharmazie* **38**: 597-600 (1983)
 41. R.K. Müller and H. Holzapfel. Parameter der flüssig/flüssig-Extraktion toxikologisch relevanter organischer Verbindungen. 4. Mitteilung: Dissoziations- und Verteilungsverhalten amphoterer organischer Elektrolyte. *Die Pharmazie* **38**: 721-728 (1983)
 42. U. Demme, U. Müller. Zur Ausbeutebestimmung toxikologisch-chemischer Analysenverfahren. *Toxichem + Krimtech* **57**: 121-125 (1990)
 43. A.D. Fraser and W. Bryan. Evaluation of the Abbott Adx and TDx serum benzodiazepines immunoassays for analysis of alprazolam. *J. Anal. Toxicol.* **15**: 63-65 (1991)
 44. W. Huang, D.E. Moody, D.M. Adrenyak and D.E. Rollins.

-
- Immunoassay detection of Nordiazepam, triazolam, Lorazepam and alprazolam in blood. *J. Anal. Toxicol.* **17**: 365-369 (1993)
45. E.R. Cairns, B.R. Dent, J.C. Ouwerkerk, and L.J. Porter. Quantitative analysis of alprazolam and triazolam in hemolysed whole blood and liver digest by GC/MS/NICI with deuterated internal standards. *J. Anal. Toxicol.* **18**: 1-6 (1994)
 46. P.M. Kabra, N.A. Mar, and L.J. Marton. Simultaneous liquid chromatographic analysis of amitriptyline, nortriptyline, imipramine, desipramine, doxepine and nordoxepine. *Clin. Chem Acta* **111**: 123-132 (1981)
 47. J.T. Cody and R. Schwarzhoff. Interpretation of methamphetamine and amphetamine enantiomer data. *J. Anal. Toxicol.* **17**: 321-326 (1993)
 48. K.S. Kalasinsky, B. Levine, M.L. Smith, J. Magluilo Jr., and T. Schaefer. Detection of amphetamine and methamphetamine by gas chromatography/fourier transform infrared (GC/FTIR) spectroscopy. *J. Anal. Toxicol.* **17**: 359-364 (1993)
 49. B.D. Paul, M.R. Past, R.M. McKinley, J.D. Foreman, L.K. McWhorter, and J.J. Snyder. Amphetamine as an artifact of methamphetamine during periodate degradation of interfering ephedrine, pseudoephedrine, and phenylpropanolamine: an improved procedure for accurate quantitation of amphetamines in urine. *J. Anal. Toxicol.* **18**: 331-336 (1994)
 50. S. Valtier and J.T. Cody. Evaluation of internal standards for the analysis of Amphetamine and methamphetamine. *J. Anal. Toxicol.* **19**: 375-380 (1995)
 51. S.N. Giorgi J.E. Meeker. A 5-year stability study of common illicit drugs in blood. *J. Anal. Toxicol.* **19**: 392-398 (1995)
 52. A. Dasgupta and J. Spies. A rapid novel derivatization of amphetamine and methamphetamine using 2,2,2-trichloroethyl chloroformate for gas chromatography electron ionization and chemical ionization mass spectrometric analysis. *Am. J. Clin. Pathol.* **109**: 527-532 (1998)
 53. J.T. Cody and S. Valtier. Detection of amphetamine and methamphetamine following administration of benzphetamine. *J. Anal. Toxicol.* **22**: 299-309 (1998)
 54. A. Poklis, J. Still, P.W. Slattum, L.F. Edinboro, J.J. Saady, and A. Constantino. Urinary excretion of d-amphetamine following oral doses in humans: implications for urine drug testing. *J. Anal. Toxicol.* **22**:

481-486 (1998)

55. D. Hensley and J.T. Cody. Simultaneous determination of amphetamine, methamphetamine, methylenedioxyamphetamine (MDA), methylenedioxy-methamphetamine (MDMA), and methylenedioxyethylamphetamine (MDEA) enantiomers by GC-MS. *J. Anal. Toxicol.* **23**: 518-523 (1999)
56. R.E. Gammans, E.H. Kerns, and W.W. Bullen. Capillary gaschromatographic mass spectrometric determination of buspirone in plasma. *J. Chromatogr.* **345**: 285-297 (1985)
57. K. Fu, R.J. Konrad, R.W. Hardy, R.M. Brissie and A.A. Robinson. An unusual multiple drug intoxication case involving citalopram. *J. Anal. Toxicol.* **24**: 648- (2000)
58. T.C. Doran. Liquid chromatographic assay for serum clonazepam. *Ther. Drug Monit.* **10**: 474-479 (1988)
59. A. El Mahjoub and C. Staub. Simultaneous determination of benzodiazepines in whole blood or serum by HPLC/DAD with a semi-micro column. *J. Pharm. Biomed. Anal.* **23**: 447-58 (2000)
60. J.E. Meeker, P.W. Herrmann, C. W. Som and P.C. Reynolds. Clozapine tissue concentrations following an apparent suicidal overdose of clozaril®. *J. Anal. Toxicol.* **16**: 54-56 (1992)
61. G.W. Hime, W.L. Hearn, S. Rose and J. Cofino. Analysis of cocaine and cocaethylene in blood and tissues by GC-NPD and GC-ion trap mass spectrometry. *J. Anal. Toxicol.* **15**: 241-245 (1991)
62. R. Martz, B. Donnelly, D. Fetterolf, L. Laswell, G.W. Hime and W.L. Hearn. The use of hair analysis to document a cocaine overdose following a sustained survival period before death. *J. Anal. Toxicol.* **15**: 279-281 (1991)
63. W.C. Brogan, III, P.M. Kemp, R.O. Bost, D.B. Glamann, R.A. Lange, and L.D. Hillis. Collection and handling of clinical blood samples to assure the accurate measurement of cocaine concentration. *J. Anal. Toxicol.* **16**: 152-154 (1992)
64. D.F. Kump, R.A. Matulka, L.E. Edinboro, A. Poklis, and M.P. Holsapple. Disposition of cocaine and norcocaine in blood and tissues of B6C3F1 mice. *J. Anal. Toxicol.* **18**: 342-345 (1994)
65. M.E. Sharp, S.M. Wallace, K.W. Hindmarsh and H.W. Peel. Monitoring saliva concentrations of methaqualone, codeine, secobarbital, diphenhydramine and diazepam after single oral dosis. *J. Anal. Toxicol.* **7**: 11-14 (1983)
66. P.M. Kemp, G.N. Sneed, C.E. George, and R.F. Distefano. Postmortem distribution of nicotine and cotinine from a case involving the simultaneous administration of multiple nicotine transdermal systems. *J. Anal. Toxicol.* **21**: 310-313 (1997)

-
67. L. v. Meyer, A. Schmoltdt, and W.R. Külpmann. Hypnotika/Seativa: Benzodiazepine. In: W.R. Külpmann (ed.), *Klinisch-toxikologische Analytik*, Wiley-VCH, Weinheim, 287 – 297, (2002)
 68. J.R. Kalin, K.M. Wood, and A.J. Lee. A possible suicide by diltiazem overdose. *J. Anal. Toxicol.* **18**: 180-182 (1994)
 69. P. Singer and A. Mozayani. An overdose fatality in a child involving disopyramide and sulindac. *J. Anal. Toxicol.* **19**: 529-530 (1995)
 70. G. de Groot, J.G. Leferink and R.A.A. Maes. Toxicological determination in biological material of doxepine by gas chromatography and some of its metabolites by mass fragmentography. *J. Anal. Toxicol.* **2**: 13 (1978)
 71. G. de Groot, R.A.A. Maes, R.C. Kelly, R.O. Bost and I. Sunshine. Four cases of fatal doxepine poisoning. *J. Anal. Toxicol.* **2**: 18 (1978)
 72. B. Fryirs, A. Woodhouse, J.L. Huang, M. Dawson, and L.E. Mather. Determination of subnanogram concentrations of fentanyl in plasma by gas chromatography-mass spectrometry: comparison with standard radioimmunoassay. *J. Chromatogr. B Biomed. Appl.* **688**: 79-85 (1997)
 73. D.T. Anderson. Two overdose fatalities involving health care professionals: postmortem tissue distribution of fentanyl. *J. Anal. Toxicol.* **21**: 93 (1997)
 74. D.T. Anderson and J.J. Muto. Duragesic® transdermal patch: postmortem tissue distribution of fentanyl in 25 cases. *J. Anal. Toxicol.* **24**: 627-634 (2000)
 75. N. Grgurinovich. A simple high-performance liquid chromatographic method for the routine measurement of flecainide in plasma. *J. Anal. Toxicol.* **12**: 38-41 (1988)
 76. C. Rogers, D. Anderson, J.K. Ribe, and L. Sathyavagiswaran. Fatal flecainide intoxication. *J. Anal. Toxicol.* **17**: 434-435 (1993)
 77. I.M. McIntyre, A. Peace, C.V. King, M.J. Lynch, and O.H. Drummer. The detection and quantitation of fluoxetine and norfluoxetine in postmortem spesies. *J. Anal. Toxicol.* **20**: 79 (1996)
 78. J.A. Crifasi, N.X. Le, and C. Long. Simultaneous identification and quantitation of fluoxetine and its metabolite, norfluoxetine, in biological samples by GC-MS. *J. Anal. Toxicol.* **21**: 415-419 (1997)
 79. B.S. Levine, S.C. Wu, B.A. Goldberger and Y.H. Caplan. Two fatalities involving haloperidol. *J. Anal. Toxicol.* **15**: 282-284 (1991)
 80. B. Levine, R.A. Jufer, and J.E. Smialek. Lamotrigine distribution in two postmortem cases. *J. Anal. Toxicol.* **24**: 635-637 (2000)
 81. P. Francom, D. Adrenyak, H.K. Lim, R.R. Bridges, R.L. Foltz, and R.T. Jones Determination of LSD in urine by capillary column gas chromatography and electron impact mass spectrometry. *J. Anal. Toxicol.* **12**: 1-8 (1988)

-
82. F. Musshoff and T. Daldrup. *Forensic Sci. Int.* **88**: 133 (1997)
 83. G.W. Kunsman, B. Levine, J.J. Kuhlman, R.L. Jones, R.O. Hughes, C.J. Fujiyama, and M.L. Smith. MDA-MDMA concentrations in urine specimens. *J. Anal. Toxicol.* **20**: 517-521 (1997)
 84. B.C. Thompson and Y.H. Caplan. A gaschromatographic method for the determination of methadone and its metabolites in biological fluids and tissues. *J. Anal. Toxicol.* **1**: 66 (1977)
 85. R.K. Lynn, R.M. Leger, W.P. Gordon, G.D. Olsen and N. Gerber. New gaschromatographic assay for the quantification of methadone. Application in human and animal studies. *J. Chromatogr.* **131**: 329-340 (1977)
 86. G. Drasch, D. Quitterer, G. Roider and L.v.Meyer. The enantioselective detection of L- and D-methadone in blood samples from living and deceased drug addicts. *Rechtsmedizin* **10**: 170-175 (2000)
 87. T.F. Simonick and V.W. Watts. Preliminary evaluation of the Abbott TDx for screening of D-methamphetamine in whole blood specimens. *J. Anal. Toxicol.* **16**: 115-118 (1992)
 88. M.A. Peat and B.S. Finkle. Determination of methaqualone and its metabolites in plasma and saliva after single oral doses. *J. Anal. Toxicol.* **4**: 114 (1980)
 89. A. Mozayani, P. Singer, and G. Jones. Distribution of metoprolol enantiomers in a fatal overdose. *J. Anal. Toxicol.* **19**: 519-521 (1995)
 90. J. Kempton, A. Manoukian, B. Levine, and J. Smialek. A mexiletine intoxication. *J. Anal. Toxicol.* **18**: 346-347 (1994)
 91. T.P. Rohrig and L.E. Harty. Postmortem distribution of mexiletine in a fatal overdose. *J. Anal. Toxicol.* **18**: 354-356 (1994)
 92. K.E. Ferslew, A.N. Hagardorn, and W.F. McCormick. Post mortem determination of the biological distribution of sulfentanil and midazolam after an acute intoxication. *J. Forensic Sci.* **34**: 249-257 (1989)
 93. B.I. Podkowik and S. Masur. Gas chromatographic determination of midazolam in low-volume plasma samples. *J. Chromatogr. B Biomed. Appl.* **681**: 405-411 (1996)
 94. D.T. Anderson, K.L. Fritz, and J.J. Muto. Distribution of mirtazapine (Remeron®) in thirteen postmortem cases. *J. Anal. Toxicol.* **23**: 544-548 (1999)
 95. P.P. Singer and G.R. Jones. An uncommon fatality due to moclobemide and paroxetine. *J. Anal. Toxicol.* **21**: 518-520 (1997)
 96. S.L. Anliker, M. Hamilton, R.J. Bopp, and M.J. Goldberg. Sensitive method for the quantitation of nortriptyline and 10-hydroxynortriptyline in

-
- human plasma by capillary gas chromatography with electron-capture detection. *J. Chromatogr.* **573**: 141-145 (1992)
97. T. Vermeulen. Distribution of paroxetine in three postmortem cases. *J. Anal. Toxicol.* **22**: 541-544 (1998)
 98. B. Levine, A. Jenkins, D. Chute, and J.E. Smialek. Perphenazine distribution in a postmortem case. *J. Anal. Toxicol.* **23**: 127-129 (1999)
 99. J.N. Miceli, D.B. Bowman and M.K. Aravind. An improved method for the quantitation of phencyclidine (PCP) in biological samples utilizing nitrogen-detection gas chromatography. *J. Anal. Toxicol.* **5**: 29-32 (1981)
 100. D.T. Anderson and K.L. Fritz. Quetiapine (Seroquel) concentrations in seven postmortem cases. *J. Anal. Toxicol.* **24**: 300-304 (2000)
 101. B.K. Logan, P.N. Friel, and G.A. Case. Analysis of sertraline (Zoloft®) and its major metabolite in postmortem specimens by gas and liquid chromatography. *J. Anal. Toxicol.* **18**: 139-142 (1994)
 102. B. Levine, A.J. Jenkins, and J.E. Smialek. Distribution of sertraline in postmortem cases. *J. Anal. Toxicol.* **18**: 272-274 (1994)
 103. D.A. Milner, M. Hall, G.G. Davis, R.M. Brissie, and C.A. Robinson. Fatal multiple drug intoxication following acute sertraline use. *J. Anal. Toxicol.* **22**: 545-548 (1998)
 104. T.G. Rosano, J.D. Hubbard, J.M. Meola and T.A. Swift. Fatal strychnine poisoning: application of gas chromatography and tandem mass spectrometry. *J. Anal. Toxicol.* **24**: 642-647 (2000)
 105. G.W. Kunsman, J.E. Manno, M.A. Przekop, B.R. Manno, K.A. Lorens, and C.M. Kunsman. Determination of temazepam and temazepam glucuronide by reversed-phase high-performance liquid chromatography. *J. Chromatogr.* **568**: 427-436 (1991)
 106. M. Kelner and D.N. Bailey. Reversed-phase liquid-chromatographic simultaneous analysis for thiopental and pentobarbital in serum. *Clin. Chem.* **29**: 1097-1100 (1983)
 107. J.R. McCutcheon. Reverse-phase HPLC determination of thioridazine and Mesoridazine in whole blood. *J. Anal. Toxicol.* **3**: 105 (1979)
 108. K.E. Goeringer, B.K. Logan, and G.D. Christian. Identification of tramadol and its metabolites in blood from drug-related deaths and drug-impaired drivers. *J. Anal. Toxicol.* **21**: 529-537 (1997)
 109. P.J. Boniface. Two cases of fatal intoxication due to tranlycypromine overdose. *J. Anal. Toxicol.* **15**: 38-40 (1991)
 110. S. Iwersen, A. Schmoltdt. One fatal and one nonfatal intoxication with tranlycypromine. Absence of amphetamines as metabolites. *J. Anal. Toxicol.* **20**: 301-304 (1996)

-
111. W.H. Anderson and M.M. Archuleta. The capillary gas chromatographic determination of trazodone in biological specimens. *J. Anal. Toxicol.* **8**: 217-219 (1984)
 112. R.E. Gammans, E.H. Kerns, W.W. Bullen, R.R. Covington, and J.W. Russell. Gas chromatographic-mass spectrometric method for trazodone and a deuterated analogue in plasma. *J. Chromatogr.* **339**: 303-312 (1985)
 113. B.P. Joynt. Triazolam blood concentrations in forensic cases in Canada. *J. Anal. Toxicol.* **17**: 171-177 (1993)
 114. R.D. Budd and D.T. Anderson. Postmortem tissue distribution of venlafaxine: six case studies. *J. Anal. Toxicol.* **21**: 93 (1997)
 115. C. Long, J. Crifasi, D. Maginn, M. Graham, and S. Teas. Comparison of analytical methods in the determination of two venlafaxin fatalities. *J. Anal. Toxicol.* **21**: 166-169 (1997)
 116. A.T. Parsons, R.M. Anthony, and J.E. Meeker. Two fatal cases of venlafaxine poisoning. *J. Anal. Toxicol.* **20**: 266-268 (1997)
 117. B. Levine, A.J. Jenkins, M. Queen, R. Jufer, and J.E. Smialek. Distribution of venlafaxine in three postmortem cases. *J. Anal. Toxicol.* **20**: 502-505 (1997)
 118. B.M. Thomson and L.K. Pannell. The analysis of verapamil in postmortem specimens by HPLC and GC. *J. Anal. Toxicol.* **5**: 105 (1981)
 119. L.F. Chan, L.H. Chhuy, and R.J. Crowley. Verapamil tissue concentrations in fatal cases. *J. Anal. Toxicol.* **11**: 171-174 (1987)
 120. B. Levine, R. Jones, K. Klette, M.L. Smith, and E. Kilbane. An intoxication involving BRONTM and verapamil. *J. Anal. Toxicol.* **17**: 381-383 (1993)
 121. J.E. Meeker, C.W. Som, E.C. Macapagal, and P.A. Benson. Zolpidem tissue concentrations in a multiple drug related death involving Ambien. *J. Anal. Toxicol.* **19**: 531-534 (1995)
 122. P.J. Boniface and S.G.G. Russell. Two cases of fatal zopiclone overdose. *J. Anal. Toxicol.* **20**: 131-133 (1996)